# Efficacy of Two Entomopathogenic Fungi, *Beauveria bassiana* and *Metarhizium anisopliae*, Isolated from Eastern Saudi Arabia against the House Fly, *Musca domestica*

# Ramy S. Yehia<sup>1,3</sup>, Essam A. Shaalan<sup>2\*</sup> and Hashem M. Al-Sheikh<sup>1</sup>

<sup>1</sup>Department of Biological Sciences, College of Science, King Faisal University, Al-Ahsa 31982, Saudi Arabia.

<sup>2</sup>Department of Zoology, Faculty of Science, Aswan University, Aswan 81528, Egypt. <sup>3</sup>Department of Botany and Microbiology, Faculty of Science, Cairo University, 12613 Giza, Egypt.

### ABSTRACT

The house fly, Musca domestica, is not only a cosmopolitan but also a medically important insect acting as vector of some diseases. Entomopathogenic fungi, particularly Beauveria bassiana and Metarhizium anisopliae, and botanical oils have shown potential as synthetic insecticides alternative for house fly control. In the present work, local isolates from both fungi as well as their mixtures with essential oils were evaluated against house fly larvae under laboratory conditions. Batches of house fly larvae (25 individuals per replicate and 4 replicates per dose of fungi) were subjected to five doses from each fungus (10<sup>1</sup>, 10<sup>3</sup>, 10<sup>5</sup>, 10<sup>7</sup> and 10<sup>9</sup> conidia/ml) in plastic cups for one min then transferred to a clean one. Both test and control cups were incubated for 7 days to determine  $LC_{50,90}$  and  $EI_{50,90}$ . Joint action of these fungi with three botanical oils (celery, ginger and sesame) as well as influence of sublethal dose from both fungi on the larval development were also evaluated. B. bassiana was more potent than M. anisopliae in both larvicidal activity and inhibition of flies' emergence. Blends from fungi and essential oils exhibited synergistic effect but fungi mixture produced antagonistic effect. The development of M. domestica larvae was affected by sublethal dose from fungi. In conclusion, M. anisopliae is more efficient than B. bassiana and could be easily mixed with essential oils to either enhance larvicidal activity or utilize in integrated pest management. Furthermore, research on field evaluation and deleterious effects of environmental conditions on fungi capacity is required.



Article Information Received 04 February 2019 Revised 14 October 2020 Accepted 20 November 2020 Available online 29 July 2021

Authors' Contribution EAS and HMA conceived and designed the project. RSY collected fungi and prepared culture. EAS performed laboratory Bioassays, analyzed data and wrote the article. HMA and RSY reviewed the manuscript.

Key words Entomopathogenic fungi, House fly larvae, Botanical oils

# INTRODUCTION

The house fly, *Musca domestica*, is a worldwide insect causing annoyance, irritation and food spoilage. Furthermore, it is important vector transmitting many pathogenic diseases including anthrax, bacillary dysentery, cholera, infantile diarrhea, tuberculosis and typhoid to both human and animals (Lecuona *et al.*, 2005; Förster *et al.*, 2009; Barin *et al.*, 2010). Accordingly, controlling this fly is crucial to avoid the previously mentioned diseases.

Synthetic insecticides were used to control the house fly (Cao *et al.*, 2006) but due to extensive use of these chemicals, insecticide resistance as well as environmental and health hazards were evolved (Bell *et al.*, 2010; Yadav, 2010). Globally, houseflies' resistance to synthetic insecticides became a big problem (Farooq and Freed, 2016) and directed the attention of the researchers to another safer alternatives exhibiting capacity in control such as entomopathogenic fungi (Zimmermann, 2007; Geden, 2012; Gul *et al.*, 2014). Among the entomopathogenic fungi, both *Beauveria bassiana* and *Metarhizium anisopliae* are the most promising insecticides alternatives against both agricultural and medically important insects in addition to some important arthropods particularly ticks and mites which are human and animals ectoparasites (Immediato *et al.*, 2015; Perinotto *et al.*, 2017).

In addition to their environmental safety and lower/ negligible mammalian toxicity, entomopathogenic fungi showed great capacity in controlling house flies (Mishra *et al.*, 2011; Khan *et al.*, 2012; Acharya *et al.*, 2015). Both *B. bassiana* (Bals.) Vuill., *M. anisopliae* (Metsch.) Sorok, were the most common fungi used in houseflies' management and produced rapid killing and high infection rates (Barson *et al.*, 1994; Kaufman *et al.*, 2005; Sharififard *et al.*, 2011). Moreover, botanical derivatives could be also introducing another alternative to synthetic insecticides.

<sup>\*</sup> Corresponding author: essamshaalan@sci.aswu.edu.eg 0030-9923/2021/0001-0001 \$ 9.00/0

Copyright 2021 Zoological Society of Pakistan

Farooq and Freed (2016) mentioned that they exhibited capacity in repelling or controlling house fly whilst several other studies shown that they could be used against all the developmental stages of the house fly (Malik *et al.*, 2007).

Recent studies have shown that botanicals could be added to entomopathogenic fungi in mixtures to synergize their potential for house fly eradication (Farooq and Freed, 2016). Ahmad *et al.* (2017) observed significant difference in larval mortality of *M. domestica* due to mixture of fungi and botanical oils except for mixtures of lower level of sub lethal doses (LC<sub>10</sub> of fungi and LC<sub>10</sub> of botanicals).

Unlike other bio insecticides, literatures revealed that data on the influence of sublethal doses of fungi on larval duration, pupal duration, pupation percentage, emergence percentage of flies and growth index are limited. The only study that was conducted by Ahmad *et al.* (2017) but for mixtures of fungi and botanical oils not for fungi alone.

Research is still in progress to find out which local isolates of the entomopathogenic fungi work effectively and can compete with synthetic insecticides. In accordance with the importance of housefly as medically and veterinary vector, the current study was carried out to investigate the efficacy of the entomopathogenic fungi *B. bassiana* and *M. anisopliae* isolated from Saudi Arabia against housefly larvae. Moreover, evaluating the effect of binary mixture from such fungi and some botanical oils as well as the effect of sublethal dose from both fungi on the development of larval stage.

# **MAERIALS AND METHODS**

# Collection, isolation, identification and purification of the entomopathogenic fungi

Both *B. bassiana* and *M. anisopliae* were the entomopathogenic fungi used in the present study. Soil samples were collected from Al-Ahsaa local livestock market during the month of August, 2017. The samples were collected in sterile glass tube for isolating the native strains of both entomopathogenic fungi in the laboratory based on morphological analysis. The fungus culture was purified by single conidia culture on potato dextrose agar (PDA) medium and subsequently sub-culturing was done according to method described by Dhingra and Sinclair (1995). Conidia were grown on PDA at 25 C in dark in standard Petri-dishes (90 mm diameter) for 10 days.

#### Spores' harvesting and suspension

The pure fungal culture was multiplied on PDA medium for 10 days. Spores were harvested by washing the dishes with pure water; subsequently the spore suspension was filtered through several layers of cheesecloth to remove mycelium. Spore concentration was determined with a

haemocytometer under light microscope and adjusted to 1X10<sup>1</sup> - 1X 10<sup>3</sup> - 1X10<sup>5</sup> - 1X10<sup>7</sup> - 1X10<sup>9</sup> spore/ ml.

#### *House fly maintenance*

The *M. domestica* flies were collected by flying insects net from livestock farms at King Faisal University, Al-Ahsaa, Eastern Saudi Arabia. Flies were transferred to insectary in Zoology department, College of Science, King Faisal University. Flies were reared in plastic cages (Collapsible insect rearing cages, Bug Dorm-1, from Bugdorm USA) measured  $30 \times 30 \times 30$  cm<sup>3</sup> under laboratory conditions. Adults were feed on 10% glucose solution, while a soluble diet made from wheat bran and milk powder (1:2) soaked in white cotton inside colorless plastic jars 300 ml capacity was for egg laying and larval development. The colony was maintained at  $28\pm2$  °C, 70  $\pm 10\%$  relative humidity and photoperiod of 14L: 10D h.

#### Larvicidal bioassays

Immersion method was used to evaluate the infectivity of two entomopathogenic fungi, *B. bassiana* and *M. anisopliae*, against larvae of *M domestica* as mentioned by Farooq and Freed (2016).

Batches of 25 newly emerged fourth instar larvae were immersed for 1 min in 1 ml of the conidial at the desired concentration  $(1 \times 10^6, 1 \times 10^7, 1 \times 10^8 \text{ spores/} \text{mL})$  of the fungi inside 20 ml plastic cups. The control group was dipped in distilled water only. Excess water was removed by Pasteur plastic pipettes. Larvae were supplied with food (dried milk, wheat barn and water in white cotton balls) and incubated at  $30 \pm 2 \circ \text{C}$ , based on preliminary experiment, for 7 days. All treatments replicated 4 times for each concentration. Mortalities were recorded after the 7<sup>th</sup> day and larvae that were immobile and did not respond to the needles counted as dead. Abbott's Formula (1925) was used for correcting observed mortalities in treatments if mortalities in control set exceeded 5% up to 10%.

#### Combined effect of fungi and essential oils

Three commercial botanical oils (celery, ginger and sesame) were used in the present results. A concentration of 10 % was freshly prepared from each oil before starting the experiments. One ml from desired oil was mixed in 10 ml distilled water and checked for one min whilst a concentration of  $1x10^3$  conidia/ml from fungi was selected for preparing the mixture. The mixtures from both oils and fungi were prepared on bases of volume to volume whereas 4 ml from both oils and fungi were mixed together in a separate glass bottle and used in the treatments.

Same procedures used for the larvicidal bioassays were followed except that each group of larvae received 2ml from the mixture rather than 1 ml. larvae were incubated at  $30 \pm 2$  °C and mortalities were recorded daily up to 7 days.

To estimate the expected synergetic effect of fungl-oil mixture, the following formula (Farenhorst *et al.*, 2010) was used:

Me = Mf + Mo (1 - Mf/100)

Where Me is expected mortality, Mf and Mo were the observed mortality percentage caused by the fungus and the oil separately. Positive Mfo - Me (observed mortality % for mixtures – expected mortality %) values were considered synergistic (Koppenhöfer and Kaya, 1998). This formula comparing mortality rates produced by fungal-oil mixture (observed) with the sum of mortalities produced by fungi and oils individually (expected).

# *Effect of sublethal dose of the fungi on the development of* M. domestica *larvae*

Methodology used for determining the larvicidal activity was adopted to investigate the influence of both fungi on some biological parameters including larval duration, pupal duration, pupation percentage, pupal mortality, adult emergence percentage and growth index. One concentration,  $4.5 \times 10^4$  conidia/ml, was used from each fungus whilst control set received distilled water. The mortalities of both larval and pupal stages as well as emerged individuals were recorded on daily bases until the emergence of the last fly or the death of the last either larva or pupa.

Larval duration was estimated as the number of days since the starting of the experiment until they all reached the pupal stage whilst the pupal duration was estimated from pupal stage to adult emergence (Martinez-Tomas *et al.*, 2009).

Growth index was calculated according to Saxena and Sumithra (1985).

Growth Index (GI) = percentage adult emergence / average developmental period (days).

The average developmental period is the sumition of larval duration and pupal duration.

#### Statistical analysis

Probit analysis was used to determine both  $LC_{50}$  and  $LC_{90}$  values whilst one-way ANOVA and the Tukey HSD post-hoc test were used for other data analysis at significance level of 5%. SPSS statistical package ver. 16 was used to perform statistical analyses.

### RESULTS

Based on  $LC_{50}$  and  $LC_{90}$  of the tested entomopathogenic fungi against *M. domestica* larvae (Table I), larvicidal activity of these fungi could be arranged in the following descending order: *B. bassiana* ( $LC_{50} = 8.6 \times 10^8$ ;  $LC_{90} = 8.9 \times 10^{12}$  conidia/ml) > *M. anisopliae* ( $LC_{50} = 1.7 \times 10^9$ ;  $LC_{90} = 6.9 \times 10^{16}$  conidia/ml). Similarly, *B. bassiana* ( $IE_{50} = 1.5 \times 10^8$ ;  $IE_{90} = 2.1 \times 10^{11}$  conidia/ml) was more potent than *M. anisopliae* ( $IE_{50} = 1.6 \times 10^9$ ;  $IE_{90} = 5.08 \times 10^{13}$  conidia/ml) in inhibiting flies emergence as has been shown in Table II.

Table I. LC<sub>50</sub> and LC<sub>90</sub> of fungi tested against newly moulted 4<sup>th</sup> instar larvae of *Musca domestica*.

Fungi	LC <sub>50</sub> Conidia/ml	LC <sub>90</sub> Conidia/ml	Chi square
Beauveria bassiana	8.6 x10 <sup>8</sup>	8.9 x10 <sup>12</sup>	113.99
Metarhizium anisopliae	1.7 x10 <sup>9</sup>	6.9 x10 <sup>16</sup>	82.72

Table II. IE<sub>50</sub> and IE<sub>90</sub> of fungi tested against newly moulted 4<sup>th</sup> instar larvae of *Musca domestica*.

Fungi	IE <sub>50</sub> Conidia/ ml	IE <sub>90</sub> Conidia/ml	Chi square
Beauveria bassiana	1.5 x10 <sup>8</sup>	2.1 x10 <sup>11</sup>	105.15
Metarhizium anisopliae	1.6 x10 <sup>9</sup>	5.08 x1013	47.07

 
 Table III. Synergistic effect of mixtures from fungi and essential oils against newly moulted 4<sup>th</sup> instar larvae of *Musca domestica*.

Mixtures/ Oils	Observed mortality Mean±SE	Expected mortality Mean±SE	Synergistic value Mean±SE
Beauveria	bassiana		
Celery	$94\pm3.83$	$72.12 \pm 6.24*$	$21.88 \pm 9.6$
Ginger	$100\pm0.0$	$69.04 \pm 7.95*$	$30.96 \pm 7.95$
Sesame	$100\pm0.0$	$74.36\pm8.74*$	$25.64 \pm 8.74$
Metarhiziu	ım anisopliae		
Celery	$96 \pm 4$	$77.08\pm6.5*$	$18.92\pm9.85$
Ginger	$96 \pm 4$	$74.36\pm8.06*$	$21.64 \pm 11.06$
Sesame	$100 \pm 0.0$	$79.76 \pm 60t.47*$	$20.24\pm6.47$

Beauveria bassiana + Metarhizium anisopliae

 $\begin{array}{cccc} 33 \pm 5.7 * & 88.68 \pm 5.26 * & -55.68 \pm 4.27 * * \\ \hline & \ \ \, \text{*, In the same column means no statistical significance (P > 0.05); **, } \\ & \ \, \text{Significantly different from all other mixtures (P < 0.05) in the same column whilst no significant difference was found among mixtures of both fungi and oils (P > 0.05). \end{array}$ 

Data of blends from fungi and essential oils against 4<sup>th</sup> instar larvae revealed synergistic effect whilst antagonistic effect was produced when both fungi mixed together (Table III). The statistical analysis Tukey HSD post-hoc test indicated that synergism produced by fungaloil blends was significantly different from synergism produced by fungal blends (F=12.377; df=6; P < 0.05). Based on synergistic value (Table III), ginger oil come in the first order in synergistic action followed by sesame oil then celery oil and all oils exhibited better synergistic action when mixed with *B. bassiana* compared with *M. anisopliae*.

Results in Table IV indicated that the sublethal dose (4.5x104 conidia/ml) of both fungi influenced the development of *M. domestica* larvae. The influence of the sublethal dose of both *B. bassiana* and *M. anisopliae* on the larval mortality percentage of *M. domestica* was not statistically significant (*F*=3.838; *df*=2; P > 0.05). Contrarily, both fungi significantly influenced larval duration (*F*=7.929; *df*=2; P < 0.05), pupal mortality (*F*=13.972; *df*=2; P < 0.05), pupal duration (*F*=12.214; *df*=2; P < 0.05), average developmental period (*F*=11.870; *df*=2; P < 0.05), adult emergence (*F*=6.499; *df*=2; P < 0.05) and growth index (*F*=10.794; *df*=2; P < 0.05) compared to control.

Table IV. Effect of sublethal dose (4.5x10<sup>4</sup> conidia/ml) on the development of *Musca domestica* larvae.

	Control	Beauveria bassiana	Metarhizium anisopliae
Larval duration	$4.25\pm0.25^{\text{ab}}$	$5.25\pm0.47^{\rm a}$	$6\pm0.0^{ab}$
Larval mortality %	1 <sup>a</sup>	$18\pm3.46^{\rm a}$	$12.75\pm6.79^{\mathrm{a}}$
Pupal duration	$6.25\pm0.25^{\text{ab}}$	$4.25\pm0.25^{\texttt{b}}$	$6\pm0.40^{ab}$
Pupal mortality %	$3\pm1.0^{ab}$	$8\pm2.3^{ab}$	$55.75 \pm 13.25^{\text{b}}$
Average develop- mental period	$10.5\pm0.28^{\rm a}$	$9.25\pm0.47^{ab}$	$12\pm0.40^{ab}$
Adult emergence %	$96\pm1.6^{ab}$	$75\pm3.4^{\mathrm{a}}$	$55\pm13.4^{ab}$
Growth index		$8.05\pm0.51^{ab}$	

<sup>a</sup>Similar letters in the same row means no statistical significance (P > 0.05); <sup>b</sup>Significantly different (P < 0.05).

### DISCUSSION

Present results revealed that the fungus *B. bassiana* is more potent larvicide ( $LC_{50} = 8.6 \times 10^8$ ;  $LC_{90} = 8.9 \times 10^{12}$  conidia/ml) against *M. domestica* larvae than *M. anisopliae* ( $LC_{50} = 1.7 \times 10^9$ ;  $LC_{90} = 6.9 \times 10^{16}$  conidia/ml). Present findings for *B. bassiana* and *M. anisopliae* isolated from Saudi Arabia are better than findings of Ibrahim *et al.* (2016) who found that the same fungi isolated from Egypt produced 7.5 and 1.25 larval mortality, respectively at a concentration of  $10^{12}$ . These results indicate that the present Saudi fungal isolate is more potent and efficient than the Egyptian one. Contrarily to present findings, Mwamburi *et al.* (2010) stated that  $LC_{50}$  of *B. bassiana* 

isolates ranged between  $10^3$ - $10^5$  conidia/ml and Mishra *et al.* (2011) indicated that *M. anisopliae* was more effective larvicide (LC<sub>50</sub> =  $4.1 \times 10^8$  conidia/ml) than *B. bassiana* (LC<sub>50</sub> =  $3.31 \times 10^9$  conidia/ml). Consequently, fungal strains may be responsible for differences in observed mortalities among house fly larvae (Weeks *et al.*, 2017). Differences in findings among these studies could be due to adopting different methods (dipping or immersing and bait techniques) for evaluating capacity of these fungi against house fly larvae/adults (Weeks *et al.*, 2017).

All mixtures produced significant larval mortalities compared with any of the agents acting alone but insignificant better synergistic action was produced by oils mixed with B. bassiana than M. anisopliae. Other similar investigations revealed the same synergistic action for blends of these fungi and some other botanical oils (Farooq and Freed, 2016; Ahmed et al., 2017). The high mortality percentage recorded after 24 h of utilizing mixtures could be due to the high synergism between these oils and fungi. Ahmad et al. (2017) reported that the synergistic action of these mixtures, entomopathogenic fungi with botanicals, could be exploited for integrated pest management (IPM) programs. In addition to their synergistic effect, it is advisable to use binary mixtures of fungal conidia with either mineral or botanical emulsified oils to avoid the deleterious effects of the environmental physical factors (Su et al., 2010; Mola and Afkari, 2012; Oliveira et al., 2018) because they protecting the conidia from such deleterious effects. In addition to their synergistic action, botanical oils can do other services for fungi. Out of seven vegetable oils, sesame showed highest effects on storage of B. bassiana conidia (Mola and Afkari, 2012). Corn oil was superior to sunflower and cotton seed oils in thermo tolerance of B. bassiana conidia (Su et al., 2010). Oliveira et al. (2018) mentioned that Oil-based formulations protect conidia from heat stress in water. They also protect entomopathogenic conidia from chemical fungicides and this advantage is being especially relevant for IPM programs whereas mycopesticides and chemicals are simultaneously sprayed (Lopes et al., 2011). Vegetable oils are also influencing the storage duration of entomopathogenic conidia (Mola and Afkari, 2012). Faria et al. (2009) reported that formulation of conidia of entomopathogenic fungi in pure (non-emulsifiable) paraffinic oil provided considerable protection from imbibitional damage.

From the present results, it is appeared that, both fungi particularly *M. anisopliae* have deleterious effects on the larvae when treated by sublethal dose nearly equal to half of the  $LC_{50}$ . Such deleterious effects included pupal mortality, the average developmental period, adult emergence and growth index. Unfortunately,

literatures revealed that studies in this aspect are very rare. Only one study mentioned the pupicdal activity of these fungi. *B. bassiana* and *M. anisopliae* caused pupal mortality percentages of 73.5 and 60.0 % at the highest concentration  $10^{12}$  Spores/ml (Ibrahim *et al.*, 2016). Compared to the present study, present findings are better particularly *M. anisopliae* that produced pupal mortality percentages of 55.75 but at lower dose by approximately two thirds which again means the strong activity of the present strain of Saudi fungal isolates.

# CONCLUSION

From these results it could be concluded that the fungus *M. anisopliae* is more efficient than *B. bassiana* and could be easily mixed with essential oils in order to either enhance their larvicidal activity or to be used in integrated pest management programs. Although these, a huge room for more research on many other disciplines are required including field evaluation and influence of environmental conditions on capacity of the entomopathogenic fungi either alone or in blends. Additionally, delivery mode in the field is another challenge. It needs more work to find convenient and more applicable mode that matches regulatory approval before commercialization.

# ACKNOWLEDGMENTS

The authors acknowledge the Deanship of Scientific Research at King Faisal University for the financial support under the annual research project (Grant No. 150148). Authors are also gratitude to Mr. Youssif Al-Gassim, Biological sciences department, College of Science, King Faisal University, Saudi Arabia for his assistance in collecting house flies and during the work.

#### Statement of conflict of interest

The authors declare that they do not have conflict of interest.

# REFERENCES

- Abbott, W.S., 1925. A method for computing the effectiveness of an insecticide. *J. econ. Ent.*, **18**: 265-267. https://doi.org/10.1093/jee/18.2.265a
- Acharya, N., Rajotte, E.G., Jenkins, N.E. and Thomas, M.B., 2015. Potential for bio-control of house flies, *Musca domestica*, using fungal bio pesticides. *Biocont. Sci. Technol.*, 25: 513-524. https://doi.org/ 10.1080/09583157.2014.992862
- Ahmed, K.W., Freed, F. and Shoukat, R.F., 2017. Efficacy of entomopathogenic fungi and botanicals

on development of *Musca domestica*. J. Ent. Zool. Stud., **5**: 593-599.

- Barin, A., Arabkhazzeli, F., Rahbari, S. and Madani, S.A., 2010. The house fly, *Musca domestica*, as a possible mechanical vector of Newcastle disease virus in the laboratory and filed. *Med. Vet. Ent.*, 24: 88-90. https://doi.org/10.1111/j.1365-2915.2009.00859.x
- Barson, G., Renn, N. and Bywater, A.F., 1994. Laboratory evaluation of six species of entomopathogenic fungi for control of house fly, *Musca domestica* L., a pest of intensive animal units. *J. Inverteb. Pathol.*, 64: 107-113. https://doi.org/10.1006/jipa.1994.1078
- Bell, H.A., Robinson, K.A. and Weaver, R.J., 2010. First report of cyromazine resistance in a population of UK house fly (*M, usca domestica*) associated with intensive livestock production. *Pest Manage. Sci.*, 66: 693-695. https://doi.org/10.1002/ps.1945
- Cao, X.M., Song, F.L., Zhao, T.Y., Dong, Y.D., Sun, C.X. and Lu, B.L., 2006. Survey of Deltamethrin resistance in houseflies (*Musca domestica*) from urban garbage dumps in Northern China. *Environ. Ent.*, **35**: 1-9. https://doi.org/10.1603/0046-225X-35.1.1
- Dhingra, O.D. and Sinclair, J.B., 1995. Basic plant pathology methods. 2<sup>nd</sup> ed. CRC Press, Boca Raton FL.
  - Farenhorst, M., Knols, B.G.J., Thomas, M.B., Howard, A.F.V., Takken, W., Rowland, M. and Guessan, R., 2010. Synergy in efficacy of fungal entomopathogens and permethrin against West African insecticide-resistant *Anopheles gambiae* mosquitoes. *PLoS One*, **5**: e12081. https://doi. org/10.1371/journal.pone.0012081
  - Faria, M., Hajek, A.E. and Wraight, S.P., 2009. Imbibitional damage in conidia of the entomopathogenic fungi *Beauveria bassiana*, *Metarhizium acridum*, and *Metarhizium anisopliae*. *Biol. Contr.*, **51**: 346-354. https://doi.org/10.1016/j. biocontrol.2009.06.012
  - Farooq, M. and Freed, S., 2016. Infectivity of housefly, *Musca domestica* (Diptera: Muscidae) to different entomopathogenic fungi. *Braz. J. Microbiol.*, 47: 807-816. https://doi.org/10.1016/j. bjm.2016.06.002
  - Förster, M., Klimpel, S. and Sievert, K., 2009. The house fly (*Musca domestica*) as a potential vector of metazoan parasites caught in a pig-pen in Germany. *Vet. Parasitol.*, **160**: 163-167. https://doi. org/10.1016/j.vetpar.2008.10.087
  - Geden, C.J., 2012. Status of biopesticides for control of house flies. *J. Biopest.*, **5**(Supplementary): 1-11.

- Gul, H.T., Saeed, S. and Khan, F.Z.A., 2014. Entomopathogenic fungi as effective insect pest management tactic: A review. *Appl. Sci. Bus. Econ.*, 1: 10-18.
- Ibrahim, A.A., Hassan, M.I., Hasaballah, A.I., Fouda, M.A. and Omar, G.M., 2016. Evaluation of some entomopathogenic fungi against larvae, pupae and adults of house fly, *Musca domestica* L. J. Nucl. Tech. appl. Sci., 41: 103-112.
- Immediato, D., Camarda, A., Iatta, R., Puttilli, M.R., Ramos, R.A.N., Di Paola, G., Giangaspero, A., Otarnto, D. and Cafarchia, C., 2015. Laboratory evaluation of a native strain of *Beauveria bassiana* for controlling *Dermanyssus gallinae* (De Geer, 1778) (Acari: Dermanyssidae). *Vet. Parasitol.*, 212: 478-482. https://doi.org/10.1016/j. vetpar.2015.07.004
- Kaufman, P.E., Reasor, C., Rutz, D.A., Ketzis, J.K. and Arends, J.J., 2005. Evaluation of *Beauveria* bassiana applications against adult house fly, *Musca domestica*, in commercial caged-layer poultry facilities in New York State. *Biol. Contr.*, **33**: 360-367. https://doi.org/10.1016/j. biocontrol.2005.03.011
- Khan, S., Guo, L.H., Maimaiti, Y., Mijit, M. and Qiu, D.W., 2012. Entomopathogenic fungi as microbial biocontrol agent. *Mol. Pl. Breed.*, **3**: 63-79. https:// doi.org/10.5376/mpb.2012.03.0007
- Koppenhöfer, A.M. and Kaya, H.K., 1998. Synergism of imidacloprid and entomopathogenic nematodes: A novel approach to white grub control in turf grass. J. econ. Ent., 91: 618-623. https://doi.org/10.1093/ jee/91.3.618
- Lecuona, R.E., Turica, M., Tarocco, F. and Crespo, D.C., 2005. Microbial control of *Musca domestica* (Diptera: Musciadae) with selected strains of *Beauveria bassiana*. J. med. Ent., 42: 332-336. https://doi.org/10.1093/jmedent/42.3.332
- Lopes, R.B., Pauli, G., Mascarin, G.M. and Faria, M., 2011. Protection of entomopathogenic conidia against chemical fungicides afforded by an oilbased formulation. *Biocont. Sci. Technol.*, **21**: 125-137. https://doi.org/10.1080/09583157.2010.53454 8
- Malik, A., Singh, N. and Satya, S., 2007. House fly (*Musca domestica*): A review of control strategies for a challenging pest. J. environ. Sci. Hlth. [B], 42: 453-469. https://doi.org/10.1080/03601230701316481
- Martinez-Tomas, S.H., Perez-Pacheco, R., Rodriguze-Hernandez, C., Ramirez-Vaverde, G. and BRuiz-Vega, J., 2009. Effects of an aqueous extract of *Azadirachta indica* on the growth of larvae and

development of pupae of *Culex quinquefasciatus*. *Afr. J. Biotechnol.*, **8**: 4245-4350.

- Mishra, S., Kumar, P., Malik, A. and Satya, S., 2011. Adulticidal and larvicidal activity of *Beauveria* bassiana and Metarhizium anisopliae against housefly, Musca domestica (Diptera: Muscidae), in laboratory and simulated field bioassays. Parasitol. Res., **108**: 148-1492. https://doi.org/10.1007/ s00436-010-2203-5
- Mola, F.L. and Afkari, R., 2012. Effects of different vegetable oils formulations on temperature tolerance and storage duration of *Beauveria bassiana* conidia. *Afr. J. Microbiol. Res.*, 6: 4707-4711. https://doi.org/10.5897/AJMR11.1372
- Mwamburi, L.A., Laing, M.D. and Miller, R.M., 2010. Laboratory screening of insecticidal activities of *Beauveria bassiana* and *Paecilomyces lilacinus* against larval and adult house fly (*Musca domestica* L.). Afr. Entomol., 18: 38-46. https://doi. org/10.4001/003.018.0106
- Oliveira, D.G.P., Lopes, R.B., Rezende, J.M. and Delalibera, I.Jr., 2018. Increased tolerance of *Beauveria bassiana* and *Metarhizium anisopale* condidia to high temperature provided by oilsbased formulations. J. Inverteb. Pathol., **151**: 151-157. https://doi.org/10.1016/j.jip.2017.11.012
- Perinotto, W.M.S., Angelo, I.C., Golo, P.S., Camargo, M.G., Quinelato, S., Sá, F.A., Coutinho Rodrigues, C.J.B., Marciano, A.F., Monteiro, C.M.O. and Bittencourt, V.R.E.P., 2017. *In vitro* pathogenicity of different *Metarhizium anisopliae* s.l. isolates in oil formulations against *Rhipicephalus microplus*. *Biocont. Sci. Technol.*, 27: 338-347. https://doi.org/ 10.1080/09583157.2017.1289151
- Saxena, S.C. and Sumithra, L., 1985. Laboratory evaluation of leaf extract of a new plant to suppress the population of malaria vector *Anopheles stephensi* Liston (Diptera: Culicidae). *Curr. Sci.*, **54**: 201-202.
- Sharififard, M., Mossadegh, M.S., Vazirianzadeh, B. and Zarei Mahmoudabadi, A., 2011. Laboratory evaluation of pathogenicity of entomopathogenic fungi, *Beauveria bassiana* (Bals.) Vuill. and *Metarhizium anisopliae* (Metch.) Sorok. to larvae and adults of the house fly, *Musca domestica* L. (Diptera: Muscidae). *Asian J. biol. Sci.*, 4: 128-137. https://doi.org/10.3923/ajbs.2011.128.137
- Su, K.J., Skinner, M. and Parker, B.L., 2010. Plant oils for improving thermotolerance of *Beauveria* bassiana. J. Microbiol. Biotechnol., 20: 1348-1350. https://doi.org/10.4014/jmb.1005.05023
- Watson, D.W., Geden, C.J., Long, S.J. and Rutz,

D.A., 1995. Efficacy of *Beauveria bassiana* for controlling the house fly and stable fly (Diptera: Musciadae). *Biol. Contr.*, **5**: 405-411. https://doi. org/10.1006/bcon.1995.1048

Weeks, E.N.I., Machtinger, E.T., Gezan, S.A., Kaufmani, P.E. and Geden, C.J., 2017. Effects of four commercial fungal formulations on mortality and sporulation in house flies (*Musca domestica*) and stable flies (*Stomoxys calcitrans*). *Med. Vet.*  *Ent.*, **31**: 15-22. https://doi.org/10.1111/mve.12201 Yadav, S.K., 2010. Pesticide applications-threat to ecosystems. *J. Hum. Ecol.*, **32**: 37-45. https://doi.or g/10.1080/09709274.2010.11906319

Zimmermann, G., 2007. Review on safety of the entomopathogenic fungus *Metarhizium anisopliae*. *Biocont. Sci. Technol.*, **17**: 879-920. https://doi. org/10.1080/09583150701593963

online